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Mechanism deconvolution of Qing Fei Pai Du decoction for treatment of Coronavirus Disease 2019 (COVID-19) by label-free integrative pharmacology assays

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ABSTRACT

Ethnopharmacological relevance: Traditional Chinese medicine (TCM) has a long history in the prevention and treatment of pandemics. The TCM formula Lung Cleansing and Detoxifying Decoction (LCDD), also known as Qing Fei Pai Du Decoction, has been demonstrated effective against Coronavirus Disease 2019 (COVID-19). Aim of the study: This work aimed to elucidate the active ingredients, targets and pathway mechanism of LCDD related to suppression of inflammatory, immunity regulation and relaxation of airway smooth muscle for the treatment of COVID-19.

Materials and methods: Mining chemical ingredients reported in LCDD, 144 compounds covering all herbs were selected and screened against inflammatory-, immunity- and respiratory-related GPCRs including GPR35, H1, CB2, B2, M3 and β 2-adrenoceptor receptor using a label-free integrative pharmacology method. Further, all active compounds were detected using liquid chromatography-tandem mass spectrometry, and an herb-compound-target network based on potency and content of compounds was constructed to elucidate the multi-target and synergistic effect.

Results: Thirteen compounds were identified as GPR35 agonists, including licochalcone B, isoliquiritigenin, etc. Licochalcone B, isoliquiritigenin and alisol A exhibited bradykinin receptor B2 antagonism activities. Attractyline and shogaol showed as a cannabinoid receptor CB2 agonist and a histamine receptor H1 antagonist, respectively. Tectorigenin and aristofone acted as muscarinic receptor M3 antagonists, while synephrine, ephedrine and pseudoephedrine were β 2-adrenoceptor agonists. Pathway deconvolution assays suggested activation of GPR35 triggered PI3K, MEK, JNK pathways and EGFR transactivation, and the activation of β 2-adrenoceptor mediated MEK and Ca²⁺. The herb-compound-target network analysis found that some compounds such as licochalcone B acted on multiple targets, and multiple components interacted with the same target such as GPR35, reflecting the synergistic mechanism of Chinese medicine. At the same time, some low-abundance compounds displayed high target activity, meaning its important role in LCDD for anti-COVID-19.

Conclusions: This study elucidates the active ingredients, targets and pathways of LCDD. This is useful for elucidating multitarget synergistic action for its clinical therapeutic efficacy.

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Table 1

ble 1 formation for lung cleansing and detoxifying decoction (LCDD). Names Uses Ephedra sinica Stapf (English: Coughs, bronchial, asthma, fever,				
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1. Introduction

Pogostemon cablin (Blanco) Benth.

(English: Pogostemonis Herba,

Chinese: Huo Xiang)

Traditional Chinese medicine (TCM) has been used in the prevention and treatment of pandemics for thousands of years. A lot of clinical experiences and effective prescriptions have been accumulated. The severe acute respiratory syndrome named Coronavirus Disease 2019

Treatment of common cold, nausea,

diarrhea, headaches and fever (Li

et al., 2011)

(COVID-19) has caused widespread infection across China and around the world since the end of 2019. In the process of fighting COVID-19, a lot of Chinese herbal preparations represented by Lung Cleansing and Detoxifying Decoction (LCDD) have been widely used in clinical practice and have shown good efficacy (Gao et al., 2020; Liu et al., 2020b; Wang et al., 2020). LCDD, also named Qing Fei Pai Du Decoction, has been widely used for the treatment of exogenous fever since 200 CE in China. It is an optimized combination of 4 classic TCM recipes, including Maxing Shigan Decoction, Wuling Powder, Xiaochaihu Decoction and Shegan Mahuang Decoction. The whole formula included 20 herbs and 1 mineral (Table 1). Maxing Shigan Decoction has been used to treat epidemics caused by viruses, such as H1N1 influenza (Wang et al., 2011) and RSV pneumonia (Jiang et al., 2015). The extracts of Glycyrrhiza uralensis Fisch. (Glycyrrhizae Radix) have beneficial effects in antiviral activities, such as human immunodeficiency virus (HIV), herpes simplex virus (HSV), SARS-CoV and etc (Yang et al., 2015). The extracts of Ephedra sinica Stapf (Ephedrae Herba) has inhibition effect on H1N1 virus (Abourashed et al., 2003). Although the components of LCDD have been shown to play a role in a variety of viral diseases, the targets, active ingredients and pathway mechanism for treatment of COVID-19 remain

Computational methods have been adopted to predict drug-target interactions of LCDD. Using the network pharmacology method, it was found that LCDD balanced immunity and eliminated inflammation by regulating several proteins co-expressed with angiotensin-converting enzyme 2 (ACE2) and signaling pathways strongly linked to the development of COVID-19 (Zhao, J. et al., 2020). Using the similar methods, other studies have shown that LCDD protected against COVID-19 via regulating anti-inflammatory, antiviral activity and cytokine storms level (Chen et al., 2020; Yang et al., 2020). Several key compounds (baicalin, glycyrrhizic acid, hesperidin, and hyperoside) and key targets (AKT1, TNF-a, IL-6, PTGS2, HMOX1, IL-10, and TP53) has been predicted (Zhao, Jing et al., 2020). However, the targets of LCDD for treatment of COVID-19 are mostly based on assumptions. Experiment based identification of the target of the main ingredient is crucial to elucidate its actual therapeutic mechanism.

The COVID-19 patients showed mild or acute respiratory syndrome accompanied by release of pro-inflammatory cytokines (Conti et al., 2020). Besides, cytokine storm is associated with clinical deterioration of COVID-19 (Huang, C. et al., 2020). Therefore, suppression of inflammatory, immunity regulation and relaxation of airway smooth muscle may have therapeutic effect on COVID-19. G protein-coupled receptors (GPCRs) are targets of approximately 33 % of the marketed human medicine, and play an important role in regulating physiological functions, such as immunity regulation, anti-inflammatory effects and respiratory regulation (Santos et al., 2017). Thus, LCDD may target a panel of GPCRs for treatment of COVID-19. The orphan GPCR GPR35 is expressed by human immune cells and its agonists play an important role in immunity regulation and anti-inflammatory activity (Quon et al., 2020). The antagonists of histamine 1 receptor (H1) are effective in suppressing inflammation (Qu et al., 2021). Cannabinoid receptor 2 (CB2) is highly expressed in multiple immune cells and its agonists show potency for the treatment of inflammatory and immune regulation (Tang et al., 2021). Bradykinin receptor 2 (B2) antagonists prevent inflammatory responses (Terzuoli et al., 2014). The muscarinic receptor 3 (M3) and β2-adrenoceptor are also involved in immunomodulatory and inflammation (Kistemaker et al., 2015; Kolmus et al., 2015). Moreover, these two receptors also play a critical role in respiratory disease, such as asthma and chronic obstructive pulmonary disease (COPD) (Patel et al., 2017; Pera and Penn, 2014). Therefore, targeting these two receptors would benefit COVID-19 patients with breathing difficulties.

In this study, we hypothesized that the active components of LCDD may function through targeting GPR35, H1, CB2, B2, M3 and β2-adrenoceptor due to their critical roles in immunomodulatory, antiinflammation and respiratory disease. These effects are highly consistent with the symptoms of COVID-19. We used a label-free integrative pharmacology method to identify active ingredients, targets and pathways of LCDD. This method uses label-free resonant waveguide grating biosensors to convert drug-induced dynamic redistribution of cellular constituents into an integrated and kinetic response, called the dynamic mass redistribution (DMR) (Schroder et al., 2011). The DMR is recorded as a shift in resonant wavelength and represents a cellular phenotypic response which covers a wide range of targets/pathways. (Grundmann and Kostenis, 2015; Ye, 2006). This assay has been extensively applied in target identification of multiple TCMs (Wang et al., 2019; Zhang et al., 2014). Herb-compound-target network was also built based on experimental data to elucidate therapeutic mechanism of LCDD. This study is useful for elucidating multitarget synergistic action of LCDD, and provides a theoretical foundation for global application in anti-COVID-19.

2. Materials and methods

2.1. Materials

Natural compounds listed in Table S1 were purchased from Shanghai Yuanye Bio-technology (Shanghai, China), Shanghai Sidend Technical Service (Shanghai, China), National Institutes for Food and Drug Control (Beijing, China), Chengdu Profa Technology Development (Chengdu, China) and Chengdu Push Bio-technology (Chengdu, China). Zaprinast, epinephrine, histamine, bradykinin, CP55940 and acetylcholine were obtained from Sigma-Aldrich (St. Louis, MO, USA). Carbachol was obtained from Tocris Bioscience Co. (St. Louis, MO, USA). Hank's balanced salt solution (HBSS), HEPES, fetal bovine serum (FBS) were from Thermo Fisher Scientific (Waltham, MA). Ham's F12K medium, Dulbecco's modified Eagle's medium (DMEM) and McCoy's 5A medium were bought from Sigma-Aldrich (St Louis, MO, USA). HPLC grade acetonitrile and methanol were purchased from Merck (Darmstadt, Germany). Water was purified from a MilliQ water system (Billerica, MA, USA). Formic acid was obtained from J&K Scientific (Beijing, China). All natural compounds were prepared in 100 mM and were dissolved in 100 % dimethyl sulfoxide (DMSO) and diluted with the assay buffer (1 × Hank's balanced salt solution (HBSS) buffer, 10 mM HEPES, pH 7.4) to the desired concentrations before DMR assays.

2.2. Cell culture

Human colorectal adenocarcinoma HT-29 cells, human epidermoid carcinoma A431 cells and human lung carcinoma A549 cells were obtained from the Type Culture Collection of the Chinese Academy of Science (Shanghai, China). HEK293-M3 cells and CHO-CB2 cells were stably transfected as previously reported (Zhou, H. et al., 2020). The cell culture medium was used as follows: 1) McCoy's 5A supplemented with 10 % FBS for HT-29 cells; 2) DMEM supplemented with 10 % FBS for A431, A549 and HEK293-M3 cells; 3) Ham's F12K medium supplemented with 10 % FBS for CHO-CB2 cells.

2.3. DMR assays

All DMR assays were performed using an Epic® BT system (Corning, NY, USA). HT-29 cells (32,000 cells per well), HEK293-M3 cells (25,000 cells per well), A431 cells (25,000 cells per well), A549 cells (20,000 cells per well) and CHO-CB2 cells (15,000 cells per well) were seeded in Epic 384-well biosensor microplates for \sim 20 h, respectively, and cells formed a monolayer on the bottom of the well in the culture medium with a confluence of \sim 95 %. Then they were directly washed with the assay buffer and maintained in 30 μL assay buffer for 1 h for DMR assay except A431 cells. The A431 cells were further incubated in the serumfree medium for 24 h. After starvation, the cells were conducted similar treatment for DMR assay.

To screen natural compounds against GPR35, H1, CB2, B2, M3 and β 2-adrenoceptor receptor in these five cell lines, a 2-min baseline was first established, followed by compound addition and DMR recording for

1 h. A 2-min baseline was then re-established, followed by adding zaprinast (2 $\mu\text{M},$ in HT-29 cells), carbachol (200 nM, in HEK293-M3 cells), epinephrine (5 nM, in A431 cells), histamine (10 $\mu\text{M},$ in A431 cells), bradykinin (500 nM, in A549 cells) and CP55940 (1 $\mu\text{M},$ in CHO-CB2) and recording DMR signals for 1 h.

For pathway deconvolution assays, HT-29 cells and A431 cells were initially treated with pathway modulators or control for 1 h. Afterwards, the baseline was re-established, followed by adding licochalcone B and ephedrine at a fixed concentration and monitoring the cellular responses induced by the compounds for 1 h.

2.4. Preparation of LCDD sample, standard solution and calibration tests

The powder sample of LCDD (20.15 mg)-gypsum fibrosum (15 g) was dissolved in 1 mL water, followed by filtration through a 0.22- μ m membrane. Stock solutions of the reference compounds were prepared in proper solvent separately at the concentration of about 0.2 mg/mL. After analysis of stock solution (injection volume 0.2 μ L), the EIC peak area was used to calculate the approximate concentration of each compound in LCDD. Then the single reference substance stock solution was diluted to the approximate concentration and reanalyzed along with LCDD (injection volume 2 μ L), using EIC peak area to calculate the exact concentration of each compound in LCDD. The sample dissolution and dilution process and the quantitative peak area were shown in Table S2.

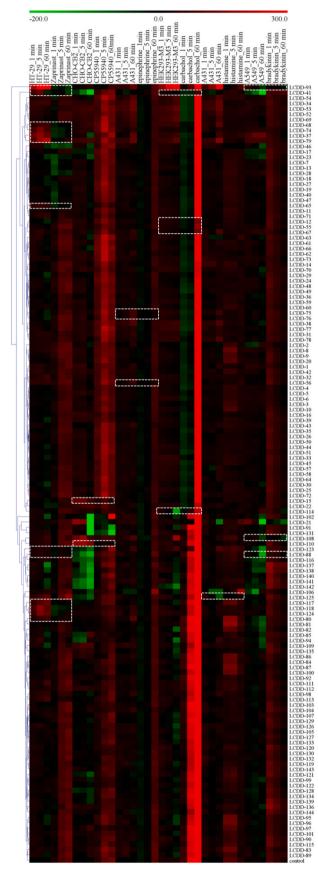
Experiments were performed on an Agilent UPLC 1290 system coupled with a high resolution quadrupole time-of-flight MS/MS 6545 system (Agilent Technologies, Santa, Clara, CA, USA), which equipped with a JetStream technology ESI interface. For chromatographic analysis, an Acquity UPLC® BEH Shield RP18 column (100 mm × 2.1 mm, 1.7 µm, Waters, Milford, MA, USA) was used. Mobile phase A was water containing 0.1 % (v/v) formic acid. Mobile phase B was acetonitrile. The linear gradient elution program was optimized as follows: 0-2 min, 100 % mobile phase A, 2-7 min, 100%-70 % mobile phase A, 7-10.5 min, 70%-40 % A, 10.5-11.5 min, 40%-10 % A, 11.5-15 min, 10 % A. The flow rate was 0.4 mL/min. The column temperature was maintained at 30 °C. The injection volume was 2 μL Mass detection was operated both in negative and positive mode. The source parameters were as follows: gas temperature 320 $^{\circ}$ C, drying gas flow rate 8 L/min, nebulizer pressure 35 psi, sheath gas temperature 350 $^{\circ}$ C, sheath gas flow rate 11 L/min, nozzle voltage (Expt) 1000 V, fragmentor voltage 135 V, skimmer voltage 65 V, collision energy 20 eV, capillary voltage 4000 V in positive mode and 3500 V in negative mode. The scan range was m/z 100-1500 for the MS scan and m/z 50-1000 for the MS/MS scan.

2.5. Networking of LCDD related to COVID-19

The recommended prescription of each herb in LCDD was collected from Diagnosis and Treatment Protocol for Novel Coronavirus Pneumonia (Trial Version 7). The content and potency value of active compounds were from HPLC-MS/MS and DMR assays, respectively. The herb attribution information corresponding to the active compounds came from the database and literatures. Import the relationship between herbs, active compounds, and targets into the Cytoscape 3.8.0, then an herb-compound-target network for LCDD was established.

2.6. Data analysis

All DMR data were acquired by EpicImager 1.20S (Corning, NY, USA) and processed by Epic Filtration v3.9.5 (Corning, NY, USA), Microsoft Excel 2010, and GraphPad Prism 6.02 (GraphPad Software Inc., San Diego, CA, USA). DMR signals were background corrected; EC_{50} and IC_{50} values were calculated by fitting the concentrations-DMR response curves with nonlinear regression.



(caption on next column)

Fig. 1. Label-free cell phenotypic heat map of 144 compounds in five cell lines including HT-29, CHO-CB2, A431, HEK293-M3 and A549 cells. This heat map was obtained using similarity analysis of the DMR signals of the compounds in these cell lines, and the DMR of the GPR35 agonist zaprinast at 2 μM in HT-29, the CB2 agonist CP55940 at 1 μM in CHO-CB2, the $\beta 2$ -adrenoceptor agonist epinephrine at 5 nM in A431, the M3 agonist carbachol at 200 nM in HEK293-M3, the H1 agonist histamine at 10 μM in A431 and the B2 agonist bradykinin at 500 nM in A549 after the pretreatment with compounds. For each compound profile, the real amplitudes at 1, 5 and 60 min after compounds treatment were used and color-coded as follows: red, positive; black, zero; green, negative. Each compound was assayed at 100 μM . All data represent mean \pm s.d. from two independent measurements, each in duplicate (n = 4). (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)

3. Results

3.1. Label-free cell phenotypic profiling of 144 compounds of LCDD

Based on chemical components in herbs of LCDD reported in literature (Zhou, Y.Y. et al., 2020), we first chose 144 compounds available in our natural product library or market. The selected compounds and their corresponding herb information were shown in Table S1. These compounds were profiled against five cell lines, including HT-29 cells (endogenously express GPR35 receptor) (Deng et al., 2011), HEK293-M3 cells, A431 cells (endogenously express β 2-adrenoceptor and H1 receptor) (Tran and Ye, 2008), A549 cells (endogenously express B2 receptor) (Izumi et al., 2018) and CHO-CB2 cells. The DMR arising from the stimulation of a specific cell line with each compound was obtained individually to determine its agonist or antagonist activity. The DMR responses of each compound were then converted into a similarity analysis (Fig. 1). According to the physiological role of these receptors, GPR35/CB2/β2 agonists or M3/H1/B2 antagonists are effective components for anti-COVID-19. In profiling, the compound is considered as an agonist if it induces significant DMR signal and blocks the receptor probe-induced DMR signal. While it is an antagonist if it does not induce obvious DMR responses and reduces the receptor probe-induced DMR signal. Herein, the agonist probe of GPR35, CB2, β2, M3, H1 and B2 was zaprinast, CP55940, epinephrine, carbachol, histamine and bradykinin, respectively.

As shown in Fig. 1, compounds led to remarkable DMR signal and attenuated zaprinast-induced DMR signal in HT-29 cells, including LCDD-93 (licochalcone B), LCDD-41 (tectorigenin), LCDD-68 (taxifolin), LCDD-74 (caffeic acid), LCDD-37 (gallic acid), LCDD-79 (isochlorogenic acid C), LCDD-65 (kynurenic acid), LCDD-123 (luteolin), LCDD-88 (isoliquiritigenin), LCDD-117 (baicalin), LCDD-118 (baicalein), LCDD-124 (scutellarin) and LCDD-80 (quercetin). The results suggested that these 13 compounds were potential GPR35 agonists. For CHO-CB2 cells, LCDD-15 (atractyline) and LCDD-110 (alisol A 24-acetate) displayed the similar characteristics, meaning that they had agonistic activity on CB2. In addition, LCDD-75 (ephedrine), LCDD-76 (pseudoephedrine) and LCDD-56 (synephrine) were possible β2-adrenoceptor agonists. LCDD-41 (tectorigenin), LCDD-12 (alismoxide), LCDD-55 (nomilin), LCDD-67 (herbacetin) and LCDD-114 (aristofone) resulted in negligible DMR signal and blocked the DMR signal induced by carbachol, indicating that these five compounds were M3 receptor antagonists. The similar behavior was observed with LCDD-125 (shogaol) for H1 receptor in A431 cells, and LCDD-93 (licochalcone B), LCDD-108 (alisol A) and LCDD-88 (isoliquiritigenin) for B2 receptor in A549 cells. Due to different patterns of active compounds detected in each receptor, the holistic view of representative active compounds for six receptors was shown in Fig. 2. Take licochalcone B as an example, it induced a positive DMR response in HT-29 cells (Fig. 2A), and the response of zaprinast was totally inhibited after cells were pretreated with licochalcone B (Fig. 2B). These results suggested that licochalcone B was a GPR35 agonist. Together, 24 potential active compounds were discovered from

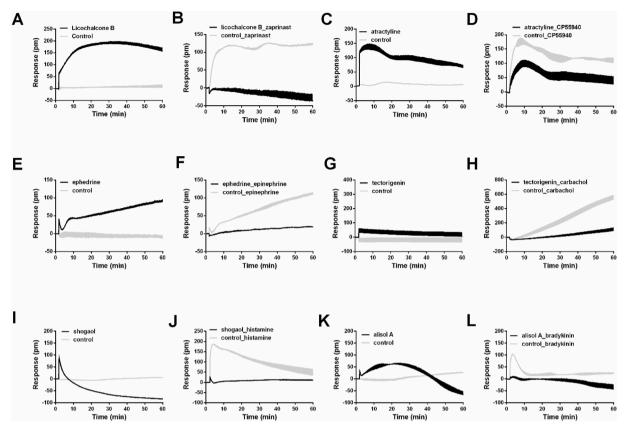


Fig. 2. Characteristic real-time DMR response. (A) Real-time DMR response of $100 \, \mu M$ licochalcone B and control in HT-29 cells. (B) Real-time DMR response of $2 \, \mu M$ zaprinast after HT-29 cells pretreated with $100 \, \mu M$ licochalcone B and control for 1 h. (C) Real-time DMR response of $100 \, \mu M$ attractyline and control in CHO-CB2 cells. (D) Real-time DMR response of $100 \, \mu M$ ephedrine and control in A431 cells. (F) Real-time DMR response of $5 \, n M$ epinephrine after A431 cells pretreated with $100 \, \mu M$ ephedrine and control for 1 h. (G) Real-time DMR response of $100 \, \mu M$ tectorigenin and control in HEK293-M3 cells. (H) Real-time DMR response of $200 \, n M$ carbachol after HEK293-M3 cells pretreated with $100 \, \mu M$ tectorigenin and control for 1 h. (I) Real-time DMR response of $100 \, \mu M$ shogaol and control in A431 cells. (J) Real-time DMR response of $100 \, \mu M$ histamine after A431 cells pretreated with $100 \, \mu M$ shogaol and control in A549 cells. (L) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (L) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (E) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (E) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (E) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (E) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (E) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (E) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (E) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (E) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells. (E) Real-time DMR response of $100 \, \mu M$ alisol A and control in A549 cells.

LCDD, suggesting that they were probably effective components in the treatment of COVID-19.

3.2. Validation of activities of potential compounds on six targets

Since the experiment in Section 3.1 was conducted using a single concentration, further multi-concentration experiments were required to confirm the activity of the candidate compounds, so as to determine if their activity were concentration dependent. Compounds that showed activity in the general screening assay (section 3.1) and caused DMR responses in a concentration-dependent manner would be determined to be agonistic or antagonistic compounds. We next evaluated the potency of 24 potential compounds on six targets. Follow-up multiple concentration tests, it was shown that licochalcone B, tectorigenin, taxifolin, isochlorogenic acid C, isoliquiritigenin, baicalin and scutellarin stimulated concentration-dependent response on HT-29 cells (Fig. 3A and Table 2). The real-time multiple concentration DMR response of licochalcone B on HT-29 cells was shown in Fig. 4A. In addition, caffeic acid (Deng and Fang, 2012), gallic acid (Deng and Fang, 2012), baicalein (Deng et al., 2012b), luteolin (Deng et al., 2012b), quercetin (Deng et al., 2012b) and kynurenic acid (Deng et al., 2012a) were reported GPR35 agonist, and their activities were also measured by DMR assay in literatures (Table 2). These results demonstrated that all potential compounds were GPR35 agonists. It is worth mentioning that licochalcone B, tectorigenin, isochlorogenic acid C and isoliquiritigenin were novel GPR35 agonist. For CHO-CB2 cells, only atractyline led to a

concentration-dependent DMR agonism response, suggesting that it was a CB2 agonist (Fig. 3B and Table 2). The real-time multiple concentration DMR response of attractyline was shown in Fig. 4B. Shogaol concentration-dependently inhibited the response of histamine in A431 cells, suggesting that it was a H1 antagonist (Fig. 3C and Table 2). The real-time multiple concentration DMR response of histamine after A431 cells were pretreated with shogaol was shown in Fig. 4C. For B2 receptor in A549 cells, licochalcone B, alisol A and isoliquiritigenin also blocked bradykinin-induced response in a concentration-dependent manner, suggesting that these three compounds were B2 antagonists (Fig. 3D and Table 2). The real-time multiple concentration DMR response of bradykinin after cells were pretreated with alisol A was shown in Fig. 4D.

For $\beta2$ -adrenoceptor in A431 cells, ephedrine, pseudoephedrine and synephrine all triggered concentration-dependent DMR response (Fig. 3E and Table 2), suggesting that these compounds were $\beta2$ -adrenoceptor agonists. Although ephedrine, pseudoephedrine and synephrine were not the first reported $\beta2$ -adrenoceptor agonists, their activities were further confirmed using DMR assay. The real-time multiple concentration DMR response of ephedrine was displayed in Fig. 4E. For M3 potential antagonists, only tectorigenin and aristofone showed concentration-dependent inhibition effects on acetylcholine-induced DMR signal, suggesting that these two compounds were M3 antagonists (Fig. 3F and Table 2). Importantly, tectorigenin and aristofone were new M3 antagonists. The real-time multiple concentration DMR response of acetylcholine after HEK293-M3 cells were pretreated with aristofone was shown in Fig. 4F. After validation, 20 compounds were

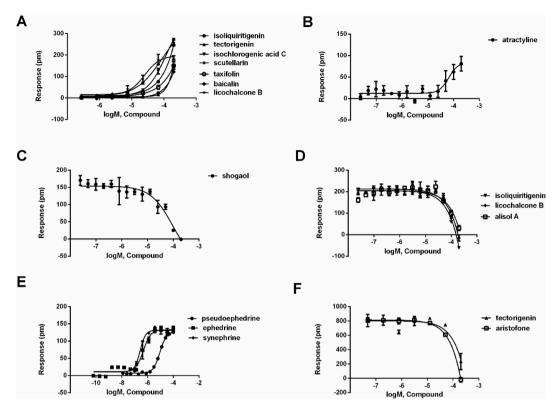


Fig. 3. Characterization of potential compounds on six targets. (A) The DMR amplitudes of licochalcone B, tectorigenin, taxifolin, isochlorogenic acid C, isoliquiritigenin, baicalin and scutellarin as a function of their concentrations in HT-29 cells. (B) The DMR amplitudes of atractyline as a function of its concentrations in CHO-CB2 cells. (C) The DMR amplitudes of histamine after A431 cells were pretreated with different concentrations of shogaol. (D) The DMR amplitudes of bradykinin after A549 cells were pretreated with different concentrations of licochalcone B, alisol A and isoliquiritigenin, respectively. (E) The DMR amplitudes of ephedrine, pseudoephedrine and synephrine as a function of their concentrations in A431 cells. (F) The DMR amplitudes of acetylcholine after HEK293-M3 cells were pretreated with different concentrations of tectorigenin and aristofone, respectively. All data represent mean \pm s.d. from two independent measurements, each in triplicate (n = 6).

identified as the active components, which had agonistic or antagonistic activities on target in a concentration-dependent manner.

3.3. Pathway deconvolution of licochalcone B in HT-29 cells and ephedrine in A431 cells

Among all active compounds, the signaling pathways of agonists were deconvoluted. Licochalcone B and ephedrine with the potency lower than 50 μM were selected as representative agonists for exploring signaling pathways of GPR35 in HT-29 cells and $\beta 2$ -adrenoceptor in A431 cells, respectively. DMR pathway deconvolution assays were used to examine the effects of the pretreatment of cells with different pathway modulators on DMR response induced by licochalcone B or ephedrine.

For the DMR response of licochalcone B in HT-29 cells, results showed that the phospholipase C (PLC) inhibitor U73122 and Ca²⁺ ATPase inhibitor thapsigargin slightly inhibited its signals (Fig. 5A), indicating that Gq-PLC-Ca²⁺ only played a small role in the DMR signal induced by licochalcone B. The PI3K inhibitor LY294002, JNK inhibitor SP600125, EGFR inhibitor AG-490 and MEK inhibitor PD98059 partially inhibited the DMR of licochalcone B. In contrast, the p38 MAPK inhibitor FHPI almost had no effect on the DMR of licochalcone B. Taken together, these results suggested that the activation of licochalcone B on GPR35 primarily triggered PI3K, JNK, MEK pathway, and EGFR transactivation.

For the DMR response of ephedrine in A431 cells, results showed that U73122 had no effect on inhibiting DMR response of ephedrine (Fig. 5B), indicating that PLC was not involved in the DMR signal induced by ephedrine. However, thapsigargin partially reduced the DMR signal, indicating that Ca^{2+} involved in the DMR signal induced by

ephedrine. LY294002 had no suppression of DMR signal of ephedrine. Similar results were observed for AG-490. FHPI and SP600125 slightly inhibited the signals of ephedrine. In contrast, PD98059 had almost 50 % inhibition of effect on the DMR. Therefore, these results suggested that the activation of ephedrine on $\beta 2$ -adrenoceptor primarily mediated MEK and Ca $^{2+}$ pathways.

3.4. Herb-compound-target-effect network for COVID-19

The content and potency value of active compounds as well as recommended prescription mass of each herb were considered in the construction of the network. To determine the content of active compounds, a HPLC-MS/MS method was established for the quantification of active compound in the LCDD samples (Fig. 6). The detailed quantitative process was shown in method section and Table S2. The quantitative results of each active compound were displayed in Table 2. A total of 17 active compounds were detected, while atractyline, shogaol and aristofone were not detected. Among these detected active compounds, the abundance of baicalin, ephedrine, pseudoephedrine, isochlorogenic acid C, scutellarin and synephrine was relatively high.

After determining the content of the active compounds, we constructed an herb-compound-target network (Fig. 7). We used the size of the rectangle to distinguish the proportion of herbs and the content of compounds in LCDD. Although Bupleuri Radix accounted for a large proportion of LCDD, we found that it contained few active compounds targeting these six GPCRs, and only baicalin and quercetin were GPR35 agonists. Ephedrae Herba occupied a medium proportion in LCDD, but we could see that there were 5 edges connecting to active compounds, which is the most among these herbs, suggesting that Ephedrae Herba is

Table 2 Potency value and content of active compounds.

NO.	Compound	Target	EC ₅₀ / IC ₅₀ (μM)	Agonist/ antagonist	Content μg/mg
1	licochalcone B	GPR35	25.12	agonist	0.000822
2	tectorigenin	GPR35	>200	agonist	0.004408
3	taxifolin	GPR35	>200	agonist	0.000486
4	isochlorogenic acid C	GPR35	114.30	agonist	0.183710
5	isoliquiritigenin	GPR35	>200	agonist	0.000073
6	baicalin	GPR35	>200	agonist	2.571457
7	scutellarin	GPR35	86.26	agonist	0.171974
8	caffeic acid	GPR35	290 a	agonist	0.0344684
9	gallic acid	GPR35	1.16 a	agonist	0.008892
10	baicalein	GPR35	10.3 a	agonist	0.003157
11	luteolin	GPR35	7.24 ^a	agonist	0.001091
12	quercetin	GPR35	8.02 a	agonist	0.003465
13	kynurenic acid	GPR35	152 ^a	agonist	0.008850
14	atractyline	CB2	65.23	agonist	undetected
15	shogaol	H1	>200	antagonist	undetected
1	licochalcone B	B2	139.70	antagonist	0.000822
16	alisol A	B2	>200	antagonist	0.001993
5	isoliquiritigenin	B2	>200	antagonist	0.000073
17	ephedrine	β2-	0.49	agonist	1.391095
		adrenoceptor			
18	pseudoephedrine	β2-	9.00	agonist	0.823086
		adrenoceptor			
19	synephrine	β2-	0.25	agonist	0.162534
		adrenoceptor			
2	tectorigenin	M3	>200	antagonist	0.0044075
20	aristofone	M3	>200	antagonist	undetected

The superscript letter a in column four indicates that the EC_{50}/IC_{50} value measured using the DMR technique comes from literatures.

very important in the treatment of COVID-19. Additionally, Scutellariae Radix, Farfarae Flos and Glycyrrhizae Radix may also play a critical role in the treatment of COVID-19, because there were also many edges connecting active compounds. We used the color of the edge between the active compound and the target to distinguish the agonistic (green) and antagonistic (red) activities and the thickness of the edge to represent the potency of the activity. We found that the content of ephedrine and pseudoephedrine both from Ephedrae Herba was relatively high, and their \(\beta 2\)-adrenoceptor agonistic activity was also strong as indicated by the size of the rectangle and thickness of the edge, which further confirmed the important role of Ephedrae Herba. Moreover, there were two edges of licochalcone B, isoliquiritigenin and tectorigeniniso connecting to targets, meaning that these compounds act on multiple targets. At the same time, there are multi-edges of compounds converging to a target, suggesting that multiple components interact with the same target such as GPR35, reflecting the synergistic mechanism of Chinese medicine. Although some compounds are very low in content in LCDD, such as licochalcone B, gallic acid, synephrine and luteolin, they may also play an important role because of their high target activity.

The SARS-CoV-2 infection inducing airway hyperresponsiveness raised the possibility the dyspnoea and respiratory failure in severe COVID-19 patients. Therefore, regulating respiration during the disease process was the crucial intervention in treating COVID-19. M3 antagonists and β 2-adrenoceptor agonists could inhibit bronchoconstriction, targeting these two receptors would benefit COVID-19 patients with breathing difficulties (Bornstein et al., 2021; De Virgiliis and Di Giovanni, 2020). We enriched the results of the respiratory disease related nodes (Fig. 8). As a result, we found that six herbs played a role in respiratory regulation, including Citri reticulatae Pericarpium, Atractylodis macrocephalae Rhizoma, Asari Radix et Rhizoma, Ephedrae Herba, Aurantii Fructus immaturus and Belamcandae Rhizoma. The

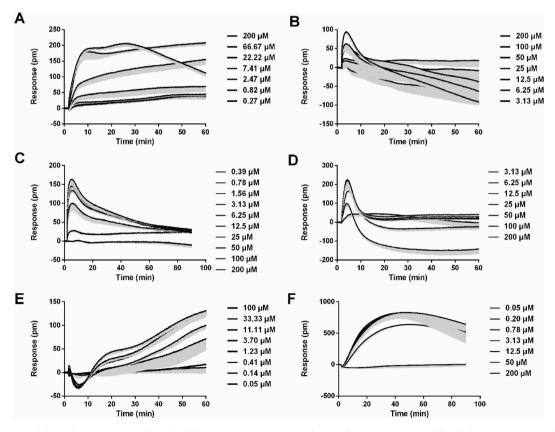


Fig. 4. Characteristics of the real-time DMR signals induced by representative compounds. (A) The DMR responses of licochalcone B in HT-29 cells. (B) The DMR responses of atractyline in CHO-CB2 cells. (C) The DMR responses of histamine after A431 cells were pretreated with shogaol. (D) The DMR responses of bradykinin after A549 cells were pretreated with alisol A. (E) The DMR responses of ephedrine in A431 cells. (F) The DMR responses of acetylcholine after HEK293-M3 cells were pretreated with aristofone. The data represent mean \pm s.d. from two independent measurements, each in triplicate (n = 6).

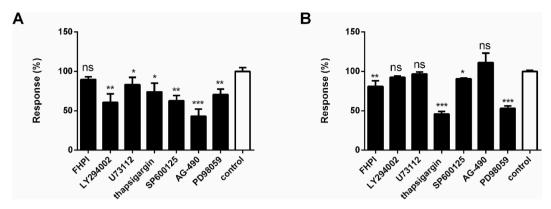
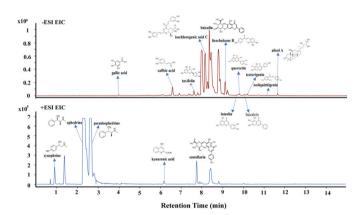


Fig. 5. Effects of pathway inhibitors FHPI (20 μ M), LY294002 (20 μ M), U73122 (20 μ M), thapsigargin (10 μ M), SP600125 (10 μ M), AG-490 (50 μ M), PD98059 (10 μ M) or control on DMR induced by licochalcone B (A), ephedrine (B). The data represent mean \pm s.d. from two independent measurements, each in triplicate (n = 6). *. P < 0.05 versus control: **. P < 0.01 versus control: ***. P < 0.001 versus control: ***. P < 0.00



 $\begin{tabular}{lll} {\bf Fig.} & {\bf 6.} & {\bf Extracted} & {\bf ion} & {\bf chromatogram} & {\bf of} & {\bf Lung} & {\bf Cleansing} & {\bf and} & {\bf Detoxifying} & {\bf Decoction}. \end{tabular}$

active compounds in LCDD here targeted all of the six GPCRs involved in suppression of inflammatory, immunity regulation and relaxation of airway smooth muscle. These results suggested that LCDD could be used as a promising formula to intervene in the pathological process of COVID-19.

4. Discussion

TCM has played an important role in the prevention and treatment of several pandemics in history. Clinical results have demonstrated the efficacy of TCMs in the treatment of COVID-19. LCDD is considered as a "universal prescription" that was recommended for COVID-19. However, its mechanism and active components remain unknown. In this study, we used a label-free integrative pharmacology method to identify the target and pathway mechanisms of active components of LCDD. As a number of COVID-19 patients showed high levels of pro-inflammatory cytokines such as IL-1, IL-6 and TNF-α, it was proposed that downregulation of inflammatory responses may improve outcome. Numerous reports have suggested a possible therapeutic role of antagonists to cytokines in the treatment of COVID-19, while some studies explored GPCRs as therapeutic targets against COVID-19. GPCRs are involved in most physiological processes in mammals, including inflammatory responses. Growing evidence demonstrated that H1 antagonists may be effective in suppressing inflammation caused by the SARS-CoV-2 infection (Qu et al., 2021). CB2 activation may have therapeutic benefit in treating COVID-19 by limiting inflammatory cytokine release (Rossi et al., 2020). B2 receptors are also possible targets for COVID-19 for regulating inflammatory responses (Kaplan and Ghebrehiwet, 2021). The SARS-CoV-2 infection induced airway hyperresponsiveness and

thus raised the possibility of dyspnoea and respiratory failure in severe COVID-19 patients (De Virgiliis and Di Giovanni, 2020). M3 antagonists could limit bronchoconstriction and vasodilation and limit the inflammatory response (Patel et al., 2017). For these reasons, we screened active component of LCDD against six GPCRs related to inflammation, immunity and respiration, including GPR35, H1, CB2, B2, M3 and β 2-adrenoceptor.

A significant finding is that licochalcone B and isoliquiritigenin both from Glycyrrhizae Radix showed dual activity against GPR35 and B2 receptors, and tectorigenin from Belamcandae Rhizoma showed dual activity against GPR35 and M3 receptor. Previous reports indicated that licochalcone B and isoliquiritigenin played an indispensable role in the potent anti-inflammatory effect of Glycyrrhiza uralensis Fisch. (Furusawa et al., 2009; Zhu et al., 2019). In addition, tectorigenin which is an effective component derived from Belamcanda chinensis, has attracted considerable interest because of its anti-inflammatory activity (Ha le et al., 2013; Pan et al., 2008). However, their targets for anti-inflammatory activity were not clear. Our findings provide a target mechanism for explaining the anti-inflammatory effect of these active components. This multi-target feature of the single compound may produce a synergistic effect in the treatment of COVID-19.

A single targeted drug may be not suitable to cure a complex disease, such as COVID-19. TCM has a good potential to complement the medical treatment for COVID-19. This may partly because of its multicomponent and multi-target effects. We explored the mechanism of LCDD against COVID-19 by pharmacology and combination strategy. A double-weighted network pharmacology, taking into account of the content weight and potency value of active compounds, was established for the first time. Previous studies have explained the treatment mechanism of LCDD through the target network model, established by predicting and collecting the targets of compounds (Chen et al., 2020; Yang et al., 2020; Zhao, J. et al., 2020). The network constructed in this study is entirely based on the results of experimental tests. It may be helpful for us to understand the actual mechanism of LCDD, especially the mechanism of GPCRs related to inflammation, immunity and respiration.

From the herb-compound-target networks, Ephedrae Herba and Glycyrrhizae Radix were considered to be very important in the treatment of COVID-19. It had been reported that Glycyrrhizae Radix was the most frequently used in prescriptions for the treatment of respiratory disease (Fu et al., 2013). The prescriptions for treatment of dyspnoea often is a combination of Herba ephedrae and Glycyrrhizae Radix (Fu et al., 2013). Interestingly, as a fundamental part of LCDD, Maxing Shigan Decoction which is recommended as a basic prescription and applied widely in the clinical treatment of COVID-19 also consists of Herba Ephedrae and Radix Glycyrrhizae. Our research also confirmed the importance of Ephedrae Herba and Glycyrrhizae Radix from the

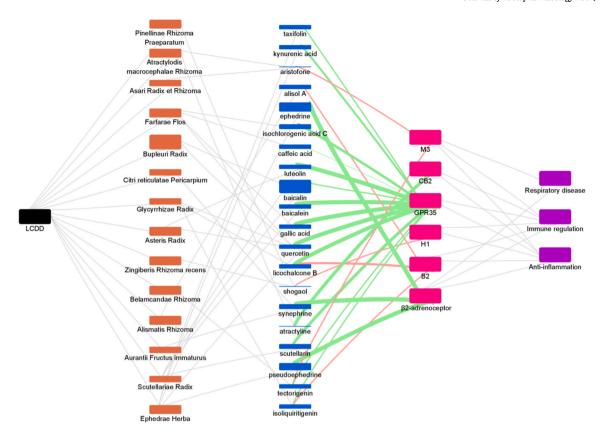


Fig. 7. The herb-compound-target network. Black rectangle: Lung Cleansing and Detoxifying Decoction (LCDD); orange rectangle: herb; blue rectangle: active compound; pink rectangle: target; purple rectangle: disease. Note: although atractyline, shogaol and aristofone were not detected by HPLC-MS/MS, they were represented by a very thin line. (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)

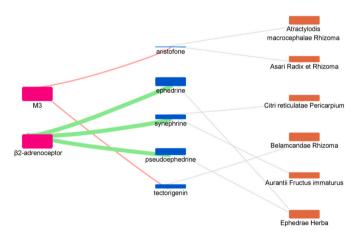


Fig. 8. The enriched herb-compound-target network of respiratory disease related nodes.

molecular mechanism level. Not only in Maxing Shigan Decoction, the active ingredients targeting GPCRs were also present in all four formulas of LCDD. As a combination recipe, LCDD completely exert the efficacy of being a multi-component and multi-target medicine. Though we discovered 20 potential active ingredients, this work is subject to a few limitations that should be noticed. First, further research would be required for the synergistic effects and metabolism of the active compounds in vivo. Second, due to the complexity of the human body and limited biotargets tested in this work, we only partially explained the therapeutic mechanism of LCDD, and more therapeutic targets should be included so as to explain the mechanism of action from a more holistic perspective. Despite the limitations, findings of this study provided new

ideas and evidence for further researches on the treatment of COVID-19 using LCDD.

The label-free DMR technology has broad signaling-pathway coverage (Rocheville et al., 2013), facilitating studies of multiple types of signaling pathways, and has the advantage in deconvoluting the signaling pathways of endogenous GPCRs. Here, we dissected the signaling pathways of GPR35 in HT-29 cells and $\beta 2$ -adrenocennptor in A431 cells. Previous studies have shown that GPR35 is a Gi and G16 coupled receptor (Taniguchi et al., 2008). The $\beta 2$ -adrenoceptor is a prototypic Gs-coupled receptor (Ferrie et al., 2014). We showed that the activation of GPR35 by licochalcone B primarily triggered PI3K, MEK, JNK pathway, EGFR transactivation, and Gq-PLC-Ca $^{2+}$ is basically not activated. The activation of ephedrine on $\beta 2$ -adrenoceptor primarily mediated MEK and Ca $^{2+}$ pathways. Currently, the identified signaling pathways are relatively upstream, and more studies are needed to establish a complete signaling pathway for the regulation of inflammation and dyspnoea caused by the SARS-CoV-2 infection.

5. Conclusions

In this study, the targets and pathways of LCDD were deconvoluted using a label-free integrative pharmacology method. 144 compounds from LCDD were profiled against six inflammatory-, immunity- and respiratory-related GPCRs. Twenty active compounds were identified for each target. Moreover, their potency and content in LCDD were determined, and the signaling pathways of licochalcone B and ephedrine were dissected. It is worth noting that a content and potency value of active compounds was both considered in the construction of the experiment-based herb-compound-target network for COVID-19. This study showed that LCDD had a protection effect on COVID-19 by regulating a panel of GPCRs and was helpful for elucidating its actual therapeutic mechanism. These results indicated that the effective treatment

of LCDD for COVID-19 may be through a holistic treatment of multi-components acting on multi-targets. Our research provides an experimental basis and research ideas for further discovery of TCM formula and botanical drug for the treatment of COVID-19.

CRediT authorship contribution statement

Fangfang Xu: Formal analysis, Writing – original draft, contributed equally to this work, designed the research; analysed the data; drafted the manuscript. All authors approved the final version of the manuscript. Tao Hou: Formal analysis, contributed equally to this work. performed the research; analysed the data; All authors approved the final version of the manuscript. Aijin Shen: performed the research; All authors approved the final version of the manuscript. Hongli Jin: performed the research; All authors approved the final version of the manuscript. Yuansheng Xiao: performed the research; All authors approved the final version of the manuscript. Wenvi Yu: performed the research; All authors approved the final version of the manuscript, performed the research; All authors approved the final version of the manuscript. Jixia Wang: Writing – original draft, designed the research; drafted the manuscript. All authors approved the final version of the manuscript. Yanfang Liu: designed the research; All authors approved the final version of the manuscript. Xinmiao Liang: designed the research; All authors approved the final version of the manuscript.

Declaration of competing interest

We wish to confirm that there are no known conflicts of interest associated with this publication and there has been no significant financial support for this work that could have influenced its outcome.

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Abbreviations

COVID-19 Coronavirus Disease 2019

TCM traditional Chinese medicine

LCDD Lung Cleansing and Detoxifying Decoction

HIV human immunodeficiency virus

HSV herpes simplex virus

ACE2 angiotensin-converting enzyme 2

GPCR G-protein coupled receptor
H1 histamine receptor 1
CB2 cannabinoid receptor 2
B2 bradykinin receptor 2
M3 muscarinic receptor 3

COPD chronic obstructive pulmonary disease

DMR dynamic mass redistribution HBSS Hank's balanced salt solution

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi. org/10.1016/j.jep.2021.114488.

References

- Abourashed, E.A., El-Alfy, A.T., Khan, I.A., Walker, L., 2003. Ephedra in perspective—a current review. Phytother Res.: PTR 17 (7), 703–712.
- Bandara, A.R., Rapior, S., Bhat, D.J., Kakumyan, P., Chamyuang, S., Xu, J.C., Hyde, K.D., 2015. Polyporus umbellatus, an edible-medicinal cultivated mushroom with

- multiple developed health-care products as Food, medicine and cosmetics: a review. Cryptog. Mycolog. 36 (1), 3–42.
- Bornstein, S.R., Voit-Bak, K., Donate, T., Rodionov, R.N., Gainetdinov, R.R., Tselmin, S., Kanczkowski, W., Muller, G.M., Achleitner, M., Wang, J., Licinio, J., Bauer, M., Young, A.H., Thuret, S., Bechmann, N., Straube, R., 2021. Chronic post-COVID-19 syndrome and chronic fatigue syndrome: is there a role for extracorporeal apheresis? Mol. Psychiatr. 1–4. https://doi.org/10.1038/s41380-021-01148-4. Advance online publication.
- Chen, J., Wang, Y.K., Gao, Y., Hu, L.S., Yang, J.W., Wang, J.R., Sun, W.J., Liang, Z.Q., Cao, Y.M., Cao, Y.B., 2020. Protection against COVID-19 injury by qingfei paidu decoction via anti-viral, anti-inflammatory activity and metabolic programming. Biomed. Pharmacother. 129, 110281.
- Chiu, C.S., Deng, J.S., Chang, H.Y., Chen, Y.C., Lee, M.M., Hou, W.C., Lee, C.Y., Huang, S. S., Huang, G.J., 2013. Antioxidant and anti-inflammatory properties of Taiwanese yam (Dioscorea japonica Thunb. var. pseudojaponica (Hayata) Yamam.) and its reference compounds. Food Chem. 141 (2), 1087–1096.
- Conti, P., Ronconi, G., Caraffa, A., Gallenga, C., Ross, R., Frydas, I., Kritas, S., 2020. Induction of pro-inflammatory cytokines (IL-1 and IL-6) and lung inflammation by Coronavirus-19 (COVI-19 or SARS-CoV-2): anti-inflammatory strategies. J. Biol. Regul. Homeost. Agents 34 (2).
- De Virgiliis, F., Di Giovanni, S., 2020. Lung innervation in the eye of a cytokine storm: neuroimmune interactions and COVID-19. Nat. Rev. Neurol. 16 (11), 645–652.
- Deng, H., Fang, Y., 2012. Anti-inflammatory gallic Acid and wedelolactone are G proteincoupled receptor-35 agonists. Pharmacology 89 (3-4), 211–219.
- Deng, H., Hu, H., Fang, Y., 2011. Tyrphostin analogs are GPR35 agonists. FEBS Lett. 585 (12), 1957–1962.
- Deng, H., Hu, H., Fang, Y., 2012a. Multiple tyrosine metabolites are GPR35 agonists. Sci. Rep. 2, 373.
- Deng, H., Hu, H., Ling, S., Ferrie, A.M., Fang, Y., 2012b. Discovery of natural phenols as G protein-coupled receptor-35 (GPR35) agonists. ACS Med. Chem. Lett. 3 (2), 165–169.
- Ferrie, A.M., Sun, H., Zaytseva, N., Fang, Y., 2014. Divergent label-free cell phenotypic pharmacology of ligands at the overexpressed beta(2)-adrenergic receptors. Sci. Rep. 4, 3828.
- Fong, W.F., Wang, C., Zhu, G.Y., Leung, C.H., Yang, M.S., Cheung, H.Y., 2007. Reversal of multidrug resistance in cancer cells by Rhizoma Alismatis extract. Phytomedicine: international journal of phytotherapy and phytopharmacology 14 (2–3), 160–165.
- international journal of phytotherapy and phytopharmacology 14 (2–3), 160–165. Fu, X.J., Song, X.X., Wei, L.B., Wang, Z.G., 2013. Study of the distribution patterns of the constituent herbs in classical Chinese medicine prescriptions treating respiratory disease by data mining methods. Chin. J. Integr. Med. 19 (8), 621–628.
- Furusawa, J., Funakoshi-Tago, M., Mashino, T., Tago, K., Inoue, H., Sonoda, Y., Kasahara, T., 2009. Glycyrrhiza inflata-derived chalcones, Licochalcone A, Licochalcone B and Licochalcone D, inhibit phosphorylation of NF-kappaB p65 in LPS signaling pathway. Int. Immunopharm. 9 (4), 499–507.
- Gao, K., Song, Y.P., Chen, H., Zhao, L.T., Ma, L., 2020. Therapeutic efficacy of Qingfei Paidu decoction combined with antiviral drugs in the treatment of corona virus disease 2019: a protocol for systematic review and meta analysis. Medicine (Baltim.) 99 (22), e20489.
- Grundmann, M., Kostenis, E., 2015. Label-free biosensor assays in GPCR screening. Methods Mol. Biol. 1272, 199–213.
- Ha le, M., Que do, T.N., Huyen do, T.T., Long, P.Q., Dat, N.T., 2013. Toxicity, analgesic and anti-inflammatory activities of tectorigenin. Immunopharmacol. Immunotoxicol. 35 (3), 336–340.
- Huang, C., Wang, Y., Li, X., Ren, L., Zhao, J., Hu, Y., Zhang, L., Fan, G., Xu, J., Gu, X., Cheng, Z., Yu, T., Xia, J., Wei, Y., Wu, W., Xie, X., Yin, W., Li, H., Liu, M., Xiao, Y., Gao, H., Guo, L., Xie, J., Wang, G., Jiang, R., Gao, Z., Jin, Q., Wang, J., Cao, B., 2020a. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 395 (10223), 497–506.
- Huang, Y.J., Hsu, N.Y., Lu, K.H., Lin, Y.E., Lin, S.H., Lu, Y.S., Liu, W.T., Chen, M.H., Sheen, L.Y., 2020b. Poria cocos water extract ameliorates the behavioral deficits induced by unpredictable chronic mild stress in rats by down-regulating inflammation. J. Ethnopharmacol. 258, 112566.
- Izumi, S., Higa-Nakamine, S., Nishi, H., Torihara, H., Uehara, A., Sugahara, K., Kakinohana, M., Yamamoto, H., 2018. Phosphorylation of epidermal growth factor receptor at serine 1047 in cultured lung alveolar epithelial cells by bradykinin B2 receptor stimulation. Pulm. Pharmacol. Therapeut. 48, 53–61.
- Jiang, L., Gao, M., Qu, F., Li, H.-l., Yu, L.-b., Rao, Y., Wang, Y.-s., Xu, G.-l., 2015.
 Pharmacokinetics of Maxing Shigan decoction in normal rats and RSV pneumonia model rats by HPLC-MS/MS. China J. Chin. Mater. Med. 40 (13), 2649–2655.
- Jung, H.J., Kim, Y.S., Shin, M.S., Kim, C.J., Kim, Y.S., 2008. Effects of armeniacae semen and amygdalin on the lipopolysaccaride-induced prostaglandin E2 synthesis and nitric oxide production in mouse BV2 microglial cells. Experimental Neurobiology 17 (2)
- Kaplan, A.P., Ghebrehiwet, B., 2021. Pathways for bradykinin formation and interrelationship with complement as a cause of edematous lung in COVID-19 patients. J. Allergy Clin. Immunol. 147 (2), 507–509.
- Kistemaker, L.E., van Os, R.P., Dethmers-Ausema, A., Bos, I.S., Hylkema, M.N., van den Berge, M., Hiemstra, P.S., Wess, J., Meurs, H., Kerstjens, H.A., Gosens, R., 2015. Muscarinic M3 receptors on structural cells regulate cigarette smoke-induced neutrophilic airway inflammation in mice. Am. J. Physiol. Lung Cell Mol. Physiol. 308 (1), L96–L103.
- Kolmus, K., Tavernier, J., Gerlo, S., 2015. beta2-Adrenergic receptors in immunity and inflammation: stressing NF-kappaB. Brain Behav. Immun. 45, 297–310.
- Li, Y.C., Xian, Y.F., Ip, S.P., Su, Z.R., Su, J.Y., He, J.J., Xie, Q.F., Lai, X.P., Lin, Z.X., 2011. Anti-inflammatory activity of patchouli alcohol isolated from Pogostemonis Herba in animal models. Fitoterapia 82 (8), 1295–1301.

- Liao, H., 2015. Effects of shengjiang (zingiberis rhizoma recens) and its processed products on nitric oxide production in macrophage RAW 264.7 cells. Evid Based Complement Alternat Med 2015, 828156.
- Lin, S., Nie, B., Yao, G., Yang, H., Ye, R., Yuan, Z., 2019. Pinellia ternata (Thunb.) Makino Preparation promotes sleep by increasing REM sleep. Nat. Prod. Res. 33 (22), 3326–3329.
- Liu, C., Wu, H., Wang, L., Luo, H., Lu, Y., Zhang, Q., Tang, L., Wang, Z., 2020a. Farfarae Flos: a review of botany, traditional uses, phytochemistry, pharmacology, and toxicology. J. Ethnopharmacol. 260, 113038.
- Liu, D., You, Y., Chen, Y., Tang, S., 2020b. Efficacy of integrative Traditional Chinese and Western medicine for the treatment of patients infected with 2019 novel coronavirus (COVID-19): a protocol for systematic review and meta analysis. Medicine (Baltim.) 99 (29), e20781.
- Liu, G.X., Xu, F., Shang, M.Y., Wang, X., Cai, S.Q., 2020c. The Relative Content and Distribution of Absorbed Volatile Organic Compounds in Rats Administered Asari Radix et Rhizoma Are Different between Powder- and Decoction-Treated Groups. Molecules 25 (19).
- Liu, J., Zhang, Q., Li, R.L., Wei, S.J., Huang, C.Y., Gao, Y.X., Pu, X.F., 2020d. The traditional uses, phytochemistry, pharmacology and toxicology of Cinnamomi ramulus: a review. J. Pharm. Pharmacol. 72 (3), 319–342.
- Ming, C.H.U., Zhengyun, C.H.U., Dandan, W., 2007. The extract of compound Radix scuteilariae on mRNA replication and IFN expression of influenza virus in mice. J. Chin. Med. Mater. 30 (1), 63–65.
- Pan, C.H., Kim, E.S., Jung, S.H., Nho, C.W., Lee, J.K., 2008. Tectorigenin inhibits IFN-gamma/LPS-induced inflammatory responses in murine macrophage RAW 264.7 cells. Arch Pharm. Res. (Seoul) 31 (11), 1447–1456.
- Patel, K.R., Bai, Y., Trieu, K.G., Barrios, J., Ai, X., 2017. Targeting acetylcholine receptor M3 prevents the progression of airway hyperreactivity in a mouse model of childhood asthma. Faseb. J. 31 (10), 4335–4346.
- Pera, T., Penn, R.B., 2014. Crosstalk between beta-2-adrenoceptor and muscarinic acetylcholine receptors in the airway. Curr. Opin. Pharmacol. 16, 72–81
- Qu, C., Fuhler, G.M., Pan, Y., 2021. Could histamine H1 receptor antagonists Be used for treating COVID-19? Int. J. Mol. Sci. 22 (11).
- Quon, T., Lin, L.C., Ganguly, A., Tobin, A.B., Milligan, G., 2020. Therapeutic opportunities and challenges in targeting the orphan G protein-coupled receptor GPR35. ACS Pharmacol Transl Sci 3 (5), 801–812.
- Rho, J., Seo, C.S., Park, H.S., Jeong, H.Y., Moon, O.S., Seo, Y.W., Son, H.Y., Won, Y.S., Kwun, H.J., 2020. Asteris Radix et Rhizoma suppresses testosterone-induced benign prostatic hyperplasia in rats by regulating apoptosis and inflammation. J. Ethnopharmacol. 255, 112779.
- Rocheville, M., Martin, J., Jerman, J., Kostenis, E., 2013. Mining the potential of label-free biosensors for seven-transmembrane receptor drug discovery. Progress in molecular biology and translational science 115, 123–142.
- Rossi, F., Tortora, C., Argenziano, M., Di Paola, A., Punzo, F., 2020. Cannabinoid receptor type 2: a possible target in SARS-CoV-2 (CoV-19) infection? Int. J. Mol. Sci. 21 (11).
- Ruqiao, L., Yueli, C., Xuelan, Z., Huifen, L., Xin, Z., Danjie, Z., Le, S., Yanxue, Z., 2020. Rhizoma Atractylodis macrocephalae: a review of photochemistry, pharmacokinetics and pharmacology. Pharmazie 75 (2), 42–55.
- Santos, R., Ursu, O., Gaulton, A., Bento, A.P., Donadi, R.S., Bologa, C.G., Karlsson, A., Al-Lazikani, B., Hersey, A., Oprea, T.I., Overington, J.P., 2017. A comprehensive map of molecular drug targets. Nat. Rev. Drug Discov. 16 (1), 19–34.
- Schroder, R., Schmidt, J., Blattermann, S., Peters, L., Janssen, N., Grundmann, M., Seemann, W., Kaufel, D., Merten, N., Drewke, C., Gomeza, J., Milligan, G., Mohr, K., Kostenis, E., 2011. Applying label-free dynamic mass redistribution technology to frame signaling of G protein-coupled receptors noninvasively in living cells. Nat. Protoc. 6 (11), 1748–1760.
- Tan, W., Li, Y., Wang, Y., Zhang, Z., Wang, T., Zhou, Q., Wang, X., 2017. Anti-coagulative and gastrointestinal motility regulative activities of Fructus Aurantii Immaturus and its effective fractions. Biomed. Pharmacother. 90, 244–252.
- Tang, Y., Wolk, B., Nolan, R., Scott, C.E., Kendall, D.A., 2021. Characterization of subtype selective cannabinoid CB2 receptor agonists as potential anti-inflammatory agents. Pharmaceuticals 14 (4).

- Taniguchi, Y., Tonai-Kachi, H., Shinjo, K., 2008. 5-Nitro-2-(3-phenylpropylamino) benzoic acid is a GPR35 agonist. Pharmacology 82 (4), 245–249.
- Terzuoli, E., Meini, S., Cucchi, P., Catalani, C., Cialdai, C., Maggi, C.A., Giachetti, A., Ziche, M., Donnini, S., 2014. Antagonism of bradykinin B2 receptor prevents inflammatory responses in human endothelial cells by quenching the NF-kB pathway activation. PloS One 9 (1), e84358.
- Tran, E., Ye, F., 2008. Duplexed label-free G protein–coupled receptor assays for high-throughput screening. J. Biomol. Screen 13 (10), 975–985.
- Wang, C., Cao, B., Liu, Q.Q., Zou, Z.Q., Liang, Z.A., Gu, L., Dong, J.P., Liang, L.R., Li, X. W., Hu, K., He, X.S., Sun, Y.H., An, Y., Yang, T., Cao, Z.X., Guo, Y.M., Wen, X.M., Wang, Y.G., Liu, Y.L., Jiang, L.D., 2011. Oseltamivir compared with the Chinese traditional therapy maxingshigan-yinqiaosan in the treatment of H1N1 influenza: a randomized trial. Ann. Intern. Med. 155 (4), 217–225.
- Wang, J., Chen, L., Qu, L., Li, K., Zhao, Y., Wang, Z., Li, Y., Zhang, X., Jin, Y., Liang, X., 2019. Isolation and bioactive evaluation of flavonoid glycosides from Lobelia chinensis Lour using two-dimensional liquid chromatography combined with labelfree cell phenotypic assays. J. Chromatogr. A 1601, 224–231.
- Wang, R.Q., Yang, S.J., Xie, C.G., Shen, Q.L., Li, M.Q., Lei, X., Li, J.K., Huang, M., 2020. Clinical observation of qingfei paidu decoction in treating COVID-19. Pharmacology and Clinics of Chinese Materia Medica 36 (1), 13–18.
- Wozniak, D., Matkowski, A., 2015. Belamcandae chinensis rhizome–a review of phytochemistry and bioactivity. Fitoterapia 107, 1–14.
- Yang, F., Dong, X., Yin, X., Wang, W., You, L., Ni, J., 2017. Radix Bupleuri: a review of traditional uses, botany, phytochemistry, pharmacology, and toxicology. BioMed Res. Int. 2017, 7597596.
- Yang, R., Liu, H., Bai, C., Wang, Y., Zhang, X., Guo, R., Wu, S., Wang, J., Leung, E., Chang, H., Li, P., Liu, T., Wang, Y., 2020. Chemical composition and pharmacological mechanism of qingfei paidu decoction and ma xing shi Gan decoction against coronavirus disease 2019 (COVID-19): in silico and experimental study. Pharmacol. Res. 157, 104820.
- Yang, R., Wang, L.-q., Yuan, B.-c., Liu, Y., 2015. The pharmacological activities of licorice. Planta Med. 81 (18), 1654–1669.
- Ye, F., 2006. Label-free cell-based assays with optical biosensors in drug discovery. Assay Drug Dev. Technol. 4 (5), 583–595.
- Yu, X., Sun, S., Guo, Y., Liu, Y., Yang, D., Li, G., Lu, S., 2018. Citri Reticulatae Pericarpium (Chenpi): botany, ethnopharmacology, phytochemistry, and pharmacology of a frequently used traditional Chinese medicine. J. Ethnopharmacol. 220, 265–282.
- Zhang, X.L., Deng, H.Y., Xiao, Y.S., Xue, X.Y., Ferrie, A.M., Tran, E., Liang, X.M., Fang, Y., 2014. Label-free cell phenotypic profiling identifies pharmacologically active compounds in two traditional Chinese medicinal plants. RSC Adv. 4 (50), 26368–26377.
- Zhao, J., Tian, S., Lu, D., Yang, J., Zeng, H., Zhang, F., Tu, D., Ge, G., Zheng, Y., Shi, T., Xu, X., Zhao, S., Yang, Y., Zhang, W., 2020a. Systems pharmacological study illustrates the immune regulation, anti-infection, anti-inflammation, and multi-organ protection mechanism of Qing-Fei-Pai-Du decoction in the treatment of COVID-19. Phytomedicine: Int. j. phytotherap. Phytopharmacol., 153315
- Zhao, J., Tian, S.S., Yang, J., Liu, J.F., Zhang, W.D., 2020b. Investigating mechanism of Qing-Fei-Pai-Du-Tang for treatment of COVID-19 by network pharmacology. Chin. Tradit. Herb. Drugs 51 (4), 829–835.
- Zhou, H., Peng, X., Hou, T., Zhao, N., Qiu, M., Zhang, X., Liang, X., 2020a. Identification of novel phytocannabinoids from Ganoderma by label-free dynamic mass redistribution assay. J. Ethnopharmacol. 246, 112218.
- Zhou, Y.Y., Gao, W.Y., Gu, X.R., Chen, Z.Q., Zhao, H.Y., Bian, B.L., Yang, L.X., Si, N., Wang, H.J., Tan, Y., 2020b. Identification and attribution of chemical constituents of Qingfei Paidu Decoction based on UHPLC-LTQ-Orbitrap-MS technology. China J. Chin. Mater. Med. 1–15.
- Zhu, X., Liu, J., Chen, S., Xue, J., Huang, S., Wang, Y., Chen, O., 2019. Isoliquiritigenin attenuates lipopolysaccharide-induced cognitive impairment through antioxidant and anti-inflammatory activity. BMC Neurosci. 20 (1), 41.